

Prevalance of Metallo Beta Lactamase Producing *Pseudomonas* Species in Clinical Specimens from S.S.G Hospital, Vadodara

Jignasha Tadvi¹, Rachna Bhavasar², Hiral Patel³, Atul Rukadikar⁴

Author Affiliation: ¹Assistant Professor ⁴Professor and Head, Department of Microbiology, Zydus Medical College & Hospital, Dahod, Gujarat 389151, India. ²Assistant Professor, Dr. M.K. Shah Medical College & Research Centre, Ahmedabad, Gujarat 382424 India. ³Tutor, GMERS Medical College, Valsad, Gujarat 396001, India.

Corresponding Author: Atul Rukadikar, Professor and Head, Department of Microbiology, Zydus Medical College & Hospital, Dahod, Gujarat 389151, India.

E-mail: atulruks@gmail.com

Received on 09.12.2018, **Accepted on** 03.01.2019

Abstract

Background: *Pseudomonas* species are the commonest pathogens causing nosocomial infections. *Pseudomonas* is basically resistant to many antibiotics and they are known to produce Extended Spectrum Beta Lactamase and Metallo beta lactamase.

AIM: To detect Metallo beta lactamase producing *Pseudomonas* spp. from clinical samples in tertiary care hospital.

Material and Methods: The study was agreed over a period of 6 months from January 2015 to June 2015. A total non repetitive of 329 *Pseudomonas* spp were isolated from unusual clinical samples like blood, pus and wound swabs, urine, body fluids, sputum, endo tracheal tube and secretions from the patients attending the hospital. Antimicrobial susceptibility test of all the isolates was performed by the disc-diffusion (Kirby Bauer disc diffusion method) according to CLSIs guidelines. All imipenem resistant isolates were tested for MBL production by Imipenem - EDTA double- disc synergy test (DDST) and Imipenem- EDTA combined disc test (CDT).

Result: Of 329 samples, majority of the *Pseudomonas* spp were isolated from Pus/Wound 173 (52.58%) followed by Blood 103 (31.31%). The isolation rate was highest from pediatrics wards 112 (34.04%) and surgical wards 112 (34.04%). Total 329 samples, 24 (7.29%) isolates were MBL producer by CDT and DDST. Majority of the MBL producing *Pseudomonas* spp. were isolated from Pus/Wound 11 (45.83%) followed by Urine 10 (41.66%). The isolation rate was highest from Surgical wards 9 (37.5%) followed by the Pediatric wards 7 (29.16%).

Conclusion: Detetion of Metallo beta lactamase isolates of *Pseudomonas* spp. will help to implement rational use of antibiotics and strictly adhere to the concept of "reserve drugs" are important to identify because it poses therapeutic problems and serious concern for infection control management.

Keywords: Metallo Beta Lactamase (MBL); *Pseudomonas* Species; Nosocomial Infections.

How to cite this article:

Jignasha Tadvi, Rachna Bhavasar, Hiral Patel et al., Prevalance of Metallo Beta Lactamase Producing *Pseudomonas* Species in Clinical Specimens from S.S.G Hospital, Vadodara. J Microbiol Relat Res. 2019;5(1):13-19.

Introduction

Pseudomonas species are the common pathogens causing nosocomial infections [1,2]. Infections caused by *Pseudomonas* are either exogenous or endogenous origin, depending on several factors such as use of immunosuppressant agents, injudious

use of antimicrobial agents, prolonged surgical procedures and inadequate instrumentations. In recent years due to liberal and empirical use of antibiotics, non fermentative gram negative bacilli have emerged as an important health care associate pathogen. They have been incriminated in infections such as septicemia, pneumonia, urinary tract

infection and surgical site infection. *Pseudomonas* is essentially resistant to many antibiotics and they are known to produce extended spectrum beta lactamase and metallo-beta lactamase. Acquired drug resistance is frequent in nosocomial isolates of *Pseudomonas* spp [1,2]. Acquired Metallo β -Lactamase (MBL) in *Pseudomonas* spp. have recently emerged as one of the most troublesome resistance mechanism because of their capability to hydrolyze all beta-lactam antibiotic including penicillins, cephalosporins and carbapenams, with the exception of Aztreonam [1,2,3,4,5]. Now a days resistance to Aztreonam producing Metallo β -Lactamase (MBL) is also revealed. Currently, there are no recommendations available from CLSI (Clinical and Laboratory Standard Institute) for the detection of MBL. Several phenotypic methods are available for MBL detection. All these methods are based on the ability of metal chelators, such as EDTA and THIOI compounds to inhibit the activity of MBL. In present study, two phenotypic methods were used for the detection of MBL producing *Pseudomonas* species which includes the Imipenem-EDTA combined disc synergy test (CDST) and Imipenem- EDTA double- disc synergy test (DDST) [3,4,5,6].

Aim and Objectives

Aim

- To determine MBL producing *Pseudomonas* spp. from clinical isolates in a tertiary care hospital setting.

Objectives

- To isolate and identify *Pseudomonas* from various clinical specimens (Blood, Body fluids, Sputum, Throat swab etc.)
- To determine antibiotic sensitivity of the isolates to various antibiotics by Kirby-Bauer disc diffusion method
- To screen for MBL producing isolates by detecting resistance to Imipenem (IPM).
- To confirm MBL production in MBL screen test positive by:
 - a. Imipenem EDTA combined disc synergy test
 - b. Imipenem EDTA double disc synergy test
- To study sensitivity and resistance pattern among isolates of *Pseudomonas* species from patients admitted in hospital.

Materials And Methods

Study Design: Cross- sectional

Study Setting: Department of Microbiology

Study Subject: The study was carried out over a period of 6 months from January 2015 to June 2015. A total non repetitive of 329 *Pseudomonas* spp were isolated from different clinical samples like blood, pus and wound swabs, urine, body fluids, sputum, endo tracheal tube and secretions from the patients attending the hospital. Antimicrobial susceptibility test of all the isolates was performed by the disc-diffusion (Kirby Baur disc diffusion method) according to CLSI guidelines. All imipenem resistant isolates were tested for MBL production by Imipenem - EDTA double- disc synergy test (DDST) and Imipenem- EDTA combined disc test (CDT).

Phenotypic method for detection of Metallo- β -Lactamases: [7,8,9,10]

Preparation of 0.5 M EDTA Solution

A 0.5 M EDTA solution was prepared by dissolving 186.1 g of disodium EDTA.2H₂O in 1,000 ml of distilled water. The pH was adjusted to 8.0 by using NaOH and was sterilized by autoclaving. The solution has to be stored at -20°C.

Combined disk test (CDT): [11,12]

The strains resistant to carbapenems were screened for MBL by CDT. Test was done for detection of metallo- β -Lactamases in the imipenem resistant isolates. An overnight liquid culture of the test isolate was adjusted to a turbidity of 0.5 McFarland standard and spread on the surface of a MHA plate. 10 μ g imipenem disk and IMP (10 μ g) + 5 μ l- 0.5 M EDTA (750 μ g) was placed on the agar.. An increase of 7mm or more in zone diameter in the presence of EDTA compared to those with IMP, tested alone was considered to be a positive test for the presence of an MBL.

Double disk synergy (DST) test: [7,8,13,14,15,16]

Test was done for detection of metallo- β -Lactamases in the imipenem resistant isolates. An overnight liquid culture of the test isolate was adjusted to a turbidity of 0.5 McFarland standard and spread on the surface of a MHA plate. A 10 μ g imipenem disk was placed on the agar. A blank disk (6 mm in diameter, Whatmann filter paper no.

1) was kept on the inner surface of the lid of the MHA plate and 10 µl of 0.5 M EDTA is added to it. This EDTA disk was then transferred to the surface of the agar and was kept 10 mm edge-to-edge apart from the imipenem disk. After incubating overnight at 37°C, the presence of an expanded growth inhibition zone between the two disks was interpreted as positive for MBL production.

Observation and Results

In this study, majority of *Pseudomonas spp.* were isolated from the age group of 1 to 10 years 91 (27.66%) and 21 to 30 years 65 (19.76%). *Pseudomonas spp.* were isolated from male patients 219 (66.57%) as compared to female patients 110 (33.43%) (Table 1).

As can be seen from Table 2, majority of the *Pseudomonas spp.* were isolated from the wound swabs and pus samples 173 (52.58%) followed by the Blood 103 (31.31%). The isolation rate of *Pseudomonas*

spp. was highest from the Surgical wards 112 (34.04%) and pediatric wards 112 (34.04%).

In the present study, of the 329 *Pseudomonas spp.* isolates, 24 (7.29%) isolates were MBL producer by CDST and DDST. While 305 (92.17%) isolates were Non- MBL producers (Table 3).

Table 1: Age and Sex distribution of isolated *Pseudomonas spp.* (n=329)

Age in Years	Male	Female	Total
< 1	11	3	14 (4.26%)
1 to 10	53	38	91 (27.66%)
11 to 20	24	12	36 (10.94%)
21 to 30	34	31	65 (19.76%)
31 to 40	28	6	34 (10.33%)
41 to 50	38	5	43 (13.07%)
51 to 60	12	9	21 (6.38%)
61 to 70	17	5	22 (6.69%)
71 to 80	2	1	3 (0.91%)
Total	219 (66.57%)	110 (33.43%)	329

Table 2: Isolation of *Pseudomonas spp.* from various clinical wards and various clinical samples.

Clinical Samples	Blood	Body fluid	Pus/ Wound	Sputum	ET/ TT	Urine	Total
Surgical	0	2	90	1	16	3	112 (34.04%)
Medical	0	0	0	4	1	1	6 (1.82%)
Pediatrics	102	0	0	0	0	10	112 (34.04%)
Orthopedic	0	0	19	0	0	0	19 (5.77%)
Obs & Gynec	0	0	2	0	0	2	4 (1.22%)
Burns	0	0	60	0	0	0	60 (18.24%)
ENT/ Eye	1	0	2	0	0	1	4 (1.22%)
Others (Ward 22,23)	0	7	0	5	0	0	12 (3.65%)
Total	103 (31.31%)	9 (2.73%)	173 (52.58%)	10 (3.04%)	17 (5.17%)	17 (5.17%)	329

Table 3: Prevalence of MBL producer and MBL non-producer of *Pseudomonas spp.*

Isolates	MBL producer N (%)	MBL non producer N (%)	Total N (%)
<i>Pseudomonas spp.</i>	24 (7.29%)	305 (92.17%)	329 (100%)

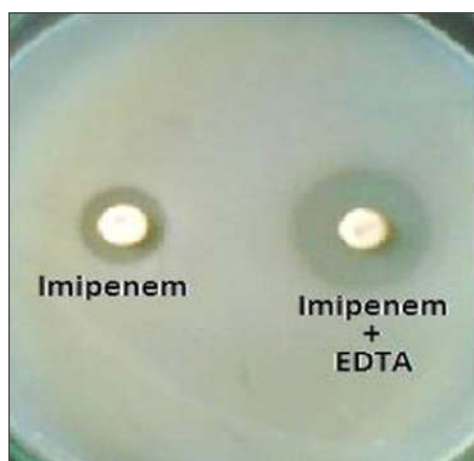


Fig 1: Combined Disk Test (CDT): positive strain shows a ≥ 7 mm zone around the Imipenem+EDTA disk.

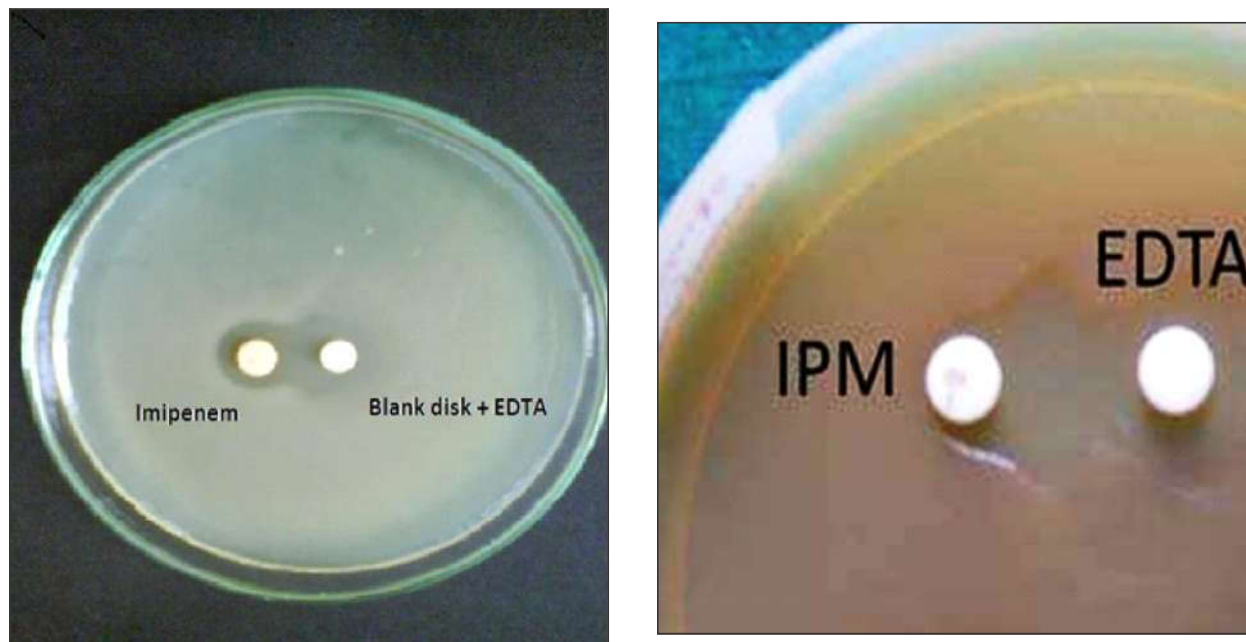


Fig 2: Double Disk Synergy Test (DDST)/ EDTA Disk Synergy Test: Positive strain shows a synergistic zone of inhibition between Imipenem and EDTA disc.

Table 4: Prevalence of MBL producing *Pseudomonas spp.* From various clinical samples and various clinical wards.

Clinical Samples	Blood	Body fluid	Pus/ Wound	Sputum	ET/ TT	Urine	Total
Surgical	0	0	7	0	1	1	9 (37.5%)
Medical	0	0	0	0	0	0	0 (0%)
Pediatrics	1	0	0	0	0	6	7 (29.16%)
Orthopedic	0	0	1	0	0	0	1 (4.17%)
Obs & Gynec	0	0	0	0	0	2	2 (8.33%)
Burns	0	0	3	0	0	0	3 (12.5%)
ENT/ Eye	0	0	0	0	0	1	1 (4.17%)
Others (Ward 22,23)	0	0	0	1	0	0	1 (4.17%)
Total	1 (4.17%)	0 (0%)	11 (45.83%)	1 (4.17%)	1 (4.17%)	10 (41.66%)	24

As can be seen from Table 4, majority of the MBL producing *Pseudomonas spp.* were isolated from Pus and wound swabs 11 (45.83%) followed by urine samples 10 (41.66%). The isolation rate was highest from surgical wards 9 (37.5%) followed by the pediatric wards 7 (29.16%).

Table 5: Antibiogram pattern of isolates of *Pseudomonas spp.* to different antibiotics are as follow:

Name of Drugs	Sensitive	Resistance
Piperacillin	278 (84.50%)	51 (15.5%)
Piperacillin + Tazobactam (100µg/ 10 µg)	312 (94.83%)	17 (5.17%)
Amikacin (30 µg)	195 (59.27%)	134 (40.73%)
Cefoperazone (75 µg)	282 (85.71%)	47 (14.29%)
Levofloxacin (5 µg)	278 (84.50%)	51 (15.5%)
Ceftazidime (30 µg)	260 (79.03%)	69 (20.97%)
Gentamicin (10 µg)	197 (59.88%)	132 (40.12%)
Imipenem (10 µg)	305 (92.71%)	24 (7.29%)

All Imipenem-resistant strains in our study showed high resistance to other antibiotics as well. High resistance was also observed to Amikacin 134 (40.73%), Gentamicin 132 (40.12%) and also Ceftazidime 69 (20.97%) and 51 (15.5%) isolates were also resistant to Piperacillin and Levofloxacin (Table 5).

Discussion

Pseudomonas spp. are the most frequent nosocomial pathogen and the infections due to these are often difficult to treat because of antibiotic resistance. Acquired drug resistance is frequent in nosocomial isolates of *Pseudomonas spp.* Acquired Metallo- β -lactamases (MBL) in *Pseudomonas spp.* have recently emerged as one of the most worrisome resistance mechanism because of their capacity

to hydrolyze all beta-lactam antibiotics including penicillins, cephalosporins and carbapenems, with the exception of aztreonam. For many years, these MBL producing isolates were restricted to Japan, but now it has disseminated worldwide. [8] In India, MBL producing *P.aeruginosa* was first reported in 2002. [8] *Pseudomonas spp.*, a virulent opportunistic pathogens which is one of the major causes of hospital acquired infection has the unique ability to infect all body systems. [17] It almost exclusively infects hospitalized patients with lowered host resistance and is the most frequent pathogens isolated from nosocomial infections in ICU.

In the present study, total 329 isolates from *Pseudomonas spp.* from different clinical samples were studied for their susceptibility or resistance to the antibiotics by (Kirby Baeur disc diffusion method) according to CLSIs guidelines.

Isolation of *Pseudomonas spp* from various clinical wards and various clinical samples.

In present study isolation of *P.seudomonas spp.* was found to be maximum from wound swabs and pus cultures 173 (52.58%) followed by blood cultures 103 (31.31%). Findings of Horieh Saderi et al. [6] (89.85%), Bashir et al. [9] (46.3%), D.E. Premalatha et al. [18] (58%) and B. Anuradha et al. [19] (39.39%) have also reported a maximum isolation from wound swabs and pus samples which corroborates well with our study. A review of surveillance data collected by the CDC National Nosocomial Infections Surveillance System

from 1986 to 1998 shows that *P. aeruginosa* was identified as the fifth most frequently isolated nosocomial pathogen, accounting for 9% of all hospital-acquired infections in the United States. *P. aeruginosa* was also the second leading cause of Nosocomial pneumonia (14 to 16%), third most common cause of urinary tract infections (7 to 11%), fourth most frequently isolated pathogen in surgical site infections (8%), and seventh leading contributor to bloodstream infections (2 to 6%).

The isolation of *Pseudomonas spp.* in our study was maximum from the Surgical wards 112 (34.04%) and pediatric wards 112 (34.04%). Rajat Rakesh et al. [20] have stated a similar finding 48% from surgical wards followed by Pediatric wards 23%.

In present study, 7.29% isolates of *P.seudomonas spp.* were found to be MBL producers phenotypically. Most of the authors [6,8,21] have mentioned similar figures. The occurrence of an MBL-positive isolates in a hospital setting poses a therapeutic problem, as well as a serious concern for infection control management [6]. In our study majority of the MBL producer *Pseudomonas spp.* were isolated from pus and wound samples 11 (45.83%) followed by the Urine samples 10 (41.66%). Bashir et al. [9] reported that the predominant source of MBL positive strains was urinary tract (27.3%) followed by wound swabs and pus (27.3%) which correlated with our study. Anil Rajput et al [22] have reported 42.9% MBL producing isolates from Wound swabs and pus samples followed by urine samples 21.4%.

Table 6: Prevalence of MBL producers among *Pseudomonas spp.*

Years of study	Place of study	Authors	MBL Producers
2005	Chennai	Hemalatha et al. [1]	87.50%
2008	Mumbai	Varaiya et al. [3]	20.80%
2008	Pakistan	S Irfan et al. [23]	100%
2008	Iran	Horieh Saderi et al. [6]	53.20%
2009	Puducherry	Noyel et al. [8]	50.00%
2010	Mumbai	Anuradha S De et al. [24]	28.57%
2011	Kashmir	Bashir et al. [9]	11.66%
2011	Tamil Nadu	John and Balagurunathan [25]	14.80%
2011	Ahmadabad	Anil Rajput [22]	12.00%
2011	Pondicherry	Umadevi S et al. [26]	74.50%
2012	Maharashtra	Simit H kumar [27]	32.04%
2013	Kolkata	Rit K et al. [21]	41%
Present study	Vadodara		7.29%

Antibiotic resistance pattern of MBL producing Pseudomonas aeruginosa isolates

All Imipenem-resistant strains in our study showed high resistance to other antibiotics as well. High resistance was also observed to Amikacin 134 (40.73%), Gentamicin 132 (40.12%) and also Ceftazidime 69 (20.97%). 51 (15.5%) isolates were also resistant to Piperacillin and Levofloxacin. This high level of resistance to Aztreonam is very alarming because it is the drug of choice for MBL producing *Pseudomonas aeruginosa*. John and Balagurunathan [25] reported 56.7% resistance to Amikacin, 100% resistance to Piperacillin, Gentamicin and Ciprofloxacin. Kumar SH et al. [27] reported that all MBL -positive isolates were resistant all antibiotics with only 6.06% of the isolates showing susceptibility of Piperacillin/Tazobactam. Irfan et al. [23] found resistance to antibiotics including third generation cephalosporin, Aminoglycoside and Quinolone. Bashir et al. [9] in his study found that the MBL producers were 100% resistant to ceftazidime, gentamicin and Tobramycin but 100% sensitive to polymyxin B. Rit et al. [21] stated that MBL producing isolates were multi drug resistant except for Colistin (100%) and for Polymyxin B (90%).

Metallo-beta-lactamase enzyme is an emerging threat and cause of concern for physician. The metal ion active site appears to decrease their susceptibility to beta lactamase inhibitors and enable them to hydrolyze broad spectrum including carbapenems. The Metallo-beta-lactamase is plasmid mediated, so the resistance can be spread among hospital pathogen and will cause problems in treating infections.

The prevalence of detect Metallo-beta-lactamase producing *Pseudomonas* spp. in our setup was 7.29%.

Conclusion

MBL (metallo- β -lactamase) positive isolates of *Pseudomonas* spp. are important to identify because it poses not only therapeutic problem, but also a serious concern for infection control management. There is also a need to emphasize on the rational use of antimicrobials and strictly adhere to the concept of "reserve drugs" to minimize the misuse of available antimicrobials. In addition, regular antimicrobial susceptibility surveillance is essential.

References

- Hemalatha V, Sekar U & Kamat V. Detection of metallo beta lactamase producing *Pseudomonas aeruginosa* in hospitalized patients. *Indian J Med Res.* 2005 Aug;122:148-52.
- Butt T, Usman M, Ahmed RN, Saif I. Emergence of Metallo- β -lactamase: The quiet before the storm? *Clinical Microbiology Reviews.* 2005;18:306-25.
- Ami Varaiya, Nikhil Kulkarni, Manasi Kulkarni, Pallavi Bhalekar & Jyotsana Dogra. Incidence of metallo beta lactamase producing *Pseudomonas aeruginosa* in ICU patients. *Indian J Med Res.* 2008 April;127:398-402.
- Debasrita Chakraborty, Saikat Basu and Satadal Das. A Study on Infections Caused By Metallo Beta Lactamase Producing Gram Negative Bacteria in Intensive Care Unit Patients. *American Journal of Infectious Diseases.* 2010;6(2):34-39.
- Gian Maria Rossolini. Acquired Metallo-b-Lactamases: An Increasing Clinical Threat. Peleg et al. on pages 1549-56.
- P Owlia, H Saderi, Z Karimi, A Rad, SM Bagher, MA Bahar. phenotypic detection of Metallo-beta-Lactamase producing *Pseudomonas aeruginosa* strains isolated from burned patients. *Iranian Journal of Pathology.* 2008;3(1):20-25.
- Lee K, Chong . Y, Shin H. B, Kim Y.A, Yong D and Yum V. Modified Hodge and EDTA- disk synergy tests to screen metallo- β - Lactamase- producing strains of *Pseudomonas* and *Acinetobacter* species. *Clin Micro Infect.* Feb 2001;7(2):88-91.
- Noyal M.J., Menezes G.A, Harish B.N., Sujatha S. & Parija S.C. Simple screening tests for detection of carbapenemases in clinical isolates of nonfermentative Gram-negative bacteria. *Indian J Med Res.* 2009 Jun;129(6):707-712.
- Bashir D, Thakor MA, Bashir AF, Bashir G, Danish Z, Shabir A and Toboli AS. Detection of metallo-b-lactamase (MBL) producing *Pseudomonas aeruginosa* at a tertiary care hospital in Kashmir. *Afr. J. Microbiol. Res.* 2011 Jan;5(2):164-172.
- Ejikeugwu Chika, Ugwu Malachy, Iroha Ifeanyi-chukwu, Eze Peter, Gugu Thaddeus, Esimone Charles. Phenotypic Detection of Metallo- β -Lactamase (MBL) Enzyme in Enugu, Southeast Nigeria. *American Journal of Biological, Chemical and Pharmaceutical Sciences.* 2014 Jun;2(2):1-6.
- Dongyeun Yong, Kyungwon Lee, Jong Hwa Yum, Hee Bong Shin, Gian Maria Rossolini and Yunsop Chong. Imipenem-EDTA Disk Method for Differentiation of Metallo- β -Lactamase-Producing Clinical Isolates of *Pseudomonas* spp. and *Acinetobacter* spp. *J Clin Microbiol.* 2002 Oct; 40(10):3798-3801.
- Seema Bose, Atindra Krishna Ghosh, Rekha Barapatre. Incidence of metallo beta lactamases

- producing *Pseudomonas aeruginosa* in burn ward of a tertiary care rural hospital. *International Journal of Biomedical Research*. 2012;3(05):233-38.
13. P.Vasundhara Devi, P.Sreenivasulu Reddy and Maria Sindhura John. Prevalence of Metallo-Lactamases Producing *Pseudomonas aeruginosa* among the Clinical isolates: A study from tertiary care hospital. *Int.J. Curr. Microbiol. App.Sci*. 2015;4(4):955-961.
 14. K. Lee, Y. S. Lim, D. Yong, J. H. Yum, and Y. Chong. Evaluation of the Hodge Test and the Imipenem-EDTA Double-Disk Synergy Test for Differentiating Metallo- β -Lactamase-Producing Isolates of *Pseudomonas* spp. and *Acinetobacter* spp. *Journal of clinical microbiology*. 2003 Oct;41(10):4623-29.
 15. Marufa Nasreen, Animesh Sarker, M. A. Malek, Md. Ansaruzzaman, Mahububur Rahman. Prevalence and Resistance Pattern of *Pseudomonas aeruginosa* Isolated from Surface Water. *Advances in Microbiology*. 2015;5:74-81.
 16. Dr. Sowmya G Shivappa, Dr. Ranjitha Shankaregowda, Dr. Raghavendra Rao M, Dr. Rajeshwari K G, Dr. Madhuri Kulkarni. Detection of Metallo-beta lactamase production in clinical isolates of Nonfermentative Gram negative bacilli. *IOSR Journal of Dental and Medical Sciences (IOSR-JDMS)*. 2015 Oct;14(10):43-48.
 17. Shenoy S, Baliga S, Saldanha D R, Prashanth H V. Antibiotic sensitivity patterns of *Pseudomonas aeruginosa* strains isolated from various clinical specimens. *Indian J Med Sci*. 2002;56(9):427-30.
 18. D E Premalatha, K C Siddesh, L H Halesh, Mallikarjun Koppad, N Prakash. Antibiotic resistance pattern of *Pseudomonas aeruginosa* strains isolated from clinical specimens in a tertiary care hospital. *Reaserch Article International Journal of Recent Trends in Science And Technology*. 2015;13(3):481-83.
 19. B. Anuradha α , Uzma Afreen σ & M. Praveena. Evaluation of Antimicrobial Susceptibility Pattern of *Pseudomonas Aeruginosa* with Special Reference to MBL Production in a Tertiary Care Hospital. *Global Journal of Medical Research: C Microbiology and Pathology*. 2014;14(7):23-28.
 20. Rajat RM, Ninama GL, Mistry K, Parmar R, Patel K, Vegad MM. Antibiotic resistance pattern in *Pseudomonas aeruginosa* species isolated at a tertiary care hospital, Ahmedabad. *National J Med. Res*. 2012 Jun;2(2):156-59.
 21. Rit K, Chakraborty B, Dey R, Chakraborty P, Naha A, Saha A. Prevalence of *Pseudomonas aeruginosa* and *Acinetobacter* spp producing metallo- β -Lactamase in a tertiary care hospital. *Jouranl of Dr. NTR University of Health Sciences*. 2013 July;2(1):18-21.
 22. Rajput A, Prajapati B, Chauhan B, Shah A, Trivedi T, Kadam M. Prevalence of Metallo-Beta Lactamase (MBL) producing *Pseudomonas aeruginosa* in a Tertiary Care Hospital. *Indian J of Basic & Applied Med Res*. 2012 Sep;1(4):304-08.
 23. S Irfan, A Zafar, D Guhar, T Ahsan, R Hasan. Metallo- β -Lactamase producing clinical isolates of *Acinetobacter* species and *Pseudomonas aeruginosa* from Intensive care unit patients of a Tertiary care hospital. *Indian J Med Microbiol*. 2008;26(3):243-45.
 24. De AS, Kumar SH and Baveja SM. Prevalence of metallo- Beta- lactamases producing *Pseudomonas aeruginosa* and *Acinetobacter* species in intensive care areas in a tertiary care hospital. *Indian J Crit Care Med*. 2010 Oct;14(4):217-219.
 25. S John, Balagurunathan R. Metallo- Beta-Lactamase producing *Pseudomonas aeruginosa* and *Acinetobacter baumannii*. *Indian J Med Microbiol*. 2011 Dec;29(3):302-304.
 26. Umadevi S, Noyal M J, Kumari K, Easow JM, Kumar S, Stephen S, Srirangaraj S, Raj S. Detection of Extended spectrum beta lactamases, AmpC beta lactamases and Metallo beta lactamases in Clinical isolates of Ceftazidime resistant *Pseudomonas aeruginosa*. *Braz. J. Microbiol*. 2011 June;42:1284-88.
 27. Kumar SH, De AS, Baveja SM, Gore MM. Prevalence and risk factor of metallo-Beta-lactamases producing *Pseudomonas aeruginosa* and *Acinetobacter* species in burns and surgical wards in a tertiary care hospital. *J Laboratory Physician*. 2012 Jan;4(1):39-42.
-